### Isolation and Characterization of Potential Bacterial Pathogens from *Cyprinus carpio* to find out the Impact of Water Pollution

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**ABSTRACT:** The most important fish pathogen is Aeromonas hydrophila (syn. A.liquifaciens, A.formicans), and this group is often referred to A.hydrophila complex. The prevalence of bacterial pathogen occurs in organically polluted waters. Predisposing risk factors include high temperature, overcrowding, organic pollution, and hypoxia. Motile aeromonads often invade skin wounds, commonly with water molds or ectoparasites. A.hydrophila is often associated with the protozoan Epistylis in causing widespread epidemic skin lesions known as red-sore disease. They are opportunistic pathogens of many immune compromised poikilotherms and homeotherms. A.hydrophila, Pseudomonas sp, Enterobacter sp., Serratia and Micrococci are some of the organisms highly abundant in the infected fishes. Among this, the maximum occurring A. hydrophila has been chosen for further microbiological and immunological studies. The direct bacterial count showed the maximum of  $37.1 \times 10^6$  CFU/ml at 48th hour whereas by the total plate count the maximum bacterial count of only  $21 \times 10^6$  CFU/ml was observed at 48th hour. The LD<sub>50</sub> value was calculated as  $3 \times 10^6$  cells for 16 gm average weight of experimental fishes. In the present study, potential bacterial pathogen like A.hydrophila was isolated and microbial characterizations were carried out to find out the impact of water pollution on the bacterial pathogenic potential.

KEYWORDS: Aeromonas hydrophila, Cyprinus carpio, water borne pathogen, fish diseases, LD<sub>50</sub>

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#### **1. INTRODUCTION**

Motile aeromonad infection (MAI) is probably the most common bacterial disease of freshwater fish. MAI has been associated with several members of the genus Aeromonas, including A.hydrophila, A.sobria, A.caviae, A.schuberri, and A.veronii. Many other species had been recently Aeromonas taxonomically identified, but only few aeromonads had been strongly documented as true fish pathogens. A. hydrophila is a free-living, ubiquitous, Gram-negative bacterium prevalent in fresh and brackish water systems [1]. A variety of freshwater and brackish water fishes such as tilapia, carp, eel, milk fish, Indian freshwater bream, Osteobrama *belangeri,* and *Plecoglossus altevelis* [2-5] have been reported to be susceptible to the aeromonad infections.

Aeromonads cause wounds [6] and systematic infections [7] in fishes. Potential virulence factors include enterotoxin, haemolysins, endotoxins, cytotoxins and proteases [8,9]. The tissues of the fish act as a barrier to prevent the entry of the hydrophobic nature of toxins [10]. Secondary invasions with motile aeromonads also characterized a wide range of other diseases, such as epizootic ulcerative syndrome (EUS) of Asian rice-field fishes, furunculosis of salmonids [11], red-sore disease of largemouth bass (*Microplexus salmodes*) [12] and many

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parasitic conditions of tropical farmed fishes, which were all showed by acute haemorrhagic septicemia and ulceration [13]. In cyprinoids, the condition may be acute, with few signs, such as large chronic ulcers [14].

*A.hydrophila* when injected intramuscularly into healthy snakehead and catfish were found to induce dermo-muscular necrotic lesions. A dose of at least 10<sup>6</sup> cells of *A.hydrophila* was required to induce EUS-like lesions in snakehead and catfish [15]. *A. hydrophila* appears to be a component of the normal bacterioflora of the rearing pond water /lake sediment and golden snails in the areas sampled. It is probable that these were the natural sources for fish infection. *A. hydrophila* involvement in superficial infections of *Catla catla* was reported in India [16].

Lactic acid bacteria are commensal organisms in the human intestine and have been found to be inhibitory towards A. hydrophila. An inhibition zone of 5 mm was observed by agar spot test. Thus Lactococcus lactis provides inhibitory effect against A. hydrophila [17]. The main objective of the present study is to find out the impact of water pollution on the pathogenecity of common carp Cyprinus carpio. Various histological samples like ulcer or lesions noted in the skin of the fish and internal organ based samples include kidney, intestine, and liver are to be microbiologically analyzed. The microbial growth pattern of the predominant bacterial species Aeromonas *hydrophila* is to be studied in detail. The LD<sub>50</sub> value of A. hydrophila, on Cyprinus carpio is planned to carry out to find the intensity of the microbial infection due to water pollution.

#### MATERIALS AND METHODS Sample Collection

In order to isolate the potential pathogenic bacteria, recently dead or moribund fish were collected from Melappalyam fish market, Palayamkottai, Tamil Nadu in sterile polyethylene bags and brought to the laboratory for further analysis.

#### Isolation of Pathogenic Bacteria

Samples were collected from the infected portions like ulcer, or lesions in the skin and from the internal organs such as kidney, intestine and liver of the diseased common carp for further enumeration and identification of pathogenic bacteria. Isolated colonies were stored in agar slants for further biochemical characterization. Cultures obtained were made free of contamination at frequent intervals by streak plating and sub culturing.

#### **Biochemical Characterization**

The standard morphological characterization criteria and biochemical tests were conducted to identify the important bacterial isolates based on Bergey's manual (1998) [18] (Table 1)

#### **Bacterial Growth Studies**

The growth pattern of selected isolates of bacteria (A. hydrophila) was studied in detail. Growth of the bacterial cells was measured by i) Direct count using Haemocytometer and ii) Total plate count method [19]. The number of cells was calculated after measuring the sample intensity or cell count at interval of 0,3,6,9,12,15,18,24,48 and 72 hours after inoculation of the cells in the fresh medium *i.e.*, each three hour interval of the incubation period cells were harvested by centrifugation. The pellet was serially diluted and total count was taken in Neubaur counting chamber. For viable count 0.1ml from the dilution was spreadplated on agar plates incubated and the colonies were counted. The count was plotted against time for estimating growth pattern.

#### Determination of LD<sub>50</sub> Value of A. hydrophila on Cyprinus carpio

To find out the LD<sub>50</sub> value of *A. hydrophila*, on *Cyprinus carpio*, eighteen hours old broth culture (at logarithmic phase) containing different loads of bacteria in physiological saline (0.85% NaCl; pH 7.2) were inoculated intraperitoneally. Six fishes were administered with a dose of  $10^3$  to  $10^8$  cells per 0.2ml. The LD<sub>50</sub> value was calculated by Reed and Muench (1938) [20]. The fish were observed carefully for visible external systems and behavioral changes. Time taken to lose the balance and the individual death was noted. The fish were considered to be dead when there was no opercular movement or response for a gentle prodding. The number of fish died was noted and their individual length and weight were measured and noted. The mortality of the challenged fish was recorded and death due to A.hydrophila was confirmed by re-isolation of organism from the liver, spleen, body fluids and intestine.

#### RESULTS

Morphological, physiological and biochemical characterization of the isolates

The occurrence of *Aeromonas hydrophila* was noticed in the diseased fresh water fish *Cyprinus carpio* from markets. According to the colony

morphology, four distinct colonies were picked up and the results are given in Table. 1.

Biochemical tests	Aeromonas hydrophila	Pseudomonas sp.	Enterobacteriaceae	Serratia marcescens
Gram staining	-	-	-	+
Motility	+	+	D	+
Kovac's oxidase test	+	_	-	D
Oxidation				
fermentationtests	+	+	-	-
Catalase tests	+	-	D	+
Cytochrome oxidase	+	+	-	-
Huge & Leifson tests	F	Ν	F	F
Starch hydrolysis	D	D	-	D
Gelatin hydrolysis	D	D	-	+
NaCl tolerance (0%)	+	D	+	+
NaCl tolerance (5%)	-	D	+	D
NaCl tolerance (7%)	-	D	D	D
Methyl Red test	+	D	+	-
Voges Proskauer	+	D	-	D
Amino acid				
decarboxylase	+	D	D	-
(Arginine)				
(Lysine)	+	D	D	D
(Ornithine)	-	D	D	+
Urease test		D	D	D
Citrate utilization test	+	D	-	+
ONPG test	D	D	+	+
0/129 sensitivity test	-	D	D	D
Growth at 5°C	-	D	D	D
Growth at 37°C	+	+	+	+

#### Table.1 Biochemical characteristics of the pathogenic isolates from Cyprinus carpio

Where, + more than 90% of identified genera show positive result;

- more than 9% of identified genera show negative result;

N- No reaction; F- Fermentation

The four chosen isolates were Gram negative except Serratia sp. They are all mesophilic, actively motile rods. Except A.hydrophila, all the other three isolates showed growth response at 5°C. At 5% and 7% concentration of NaCl except A.hydrophila all other isolates showed positive growth response. In the Indole test A. hydrophila and Serratia sp showed positive results whereas Pseudomonas sp and Enterobacter sp showed negative result. Except Serratia sp, all were Methyl red positive; except Enterobacter all were VP (Voges Proskauer) positive and citrate positive.

Amino acid decarboxylase of arginine was found to be positive for all three isolates except

*Serratia*. Lysine decarboxylase was positive for all isolates, and ornithine decarboxylase was positive for three isolates except *A.hydrophila*. All the four isolates showed ONPG positive. Except *Enterobacter* sp, all other three isolates showed positive for starch and gelatin hydrolysis. Based on the morphological, physiological and biochemical characteristics of the four isolates and on comparison with Bergey's manual (1998) [18], the identification of the bacterial isolates was confirmed. (Table.1). The four isolates were confirmed as *A.hydrophila, Pseudomonas* sp., *Enterobacter* sp. and *Serratia* sp.

D -11-89% show positive result;

## Percentage composition of bacterial genera from diseased fish Cyprinus carpio

From the isolated samples from liver, kidney, intestine and body fluid, *A.hydrophila* was found to be higher in body fluid (46%) and liver (44%). In the intestine, both *A.hvdrophila* and Enterobacter sp. were found almost in equal load (29%). But in the kidney sample, Pseudomonas sp. was found to be highest in maximum load (33%). Figure 1 to 4 shows that A.hydrophila count was found to be maximum except the kidney sample which was recorded second largest microbial count. The other microorganisms observed in the diseased histological samples of the fish Cyprinus carpio include, Pseudomonas sp, Enterobacter sp., Serratia, Micrococci, etc. The maximum microbial count of A. hydrophila was recorded during the studies. It has been chosen for further microbiological and immunological studies.

#### **Bacterial growth studies**

The viability of A.hydrophila isolates was counter-checked by growth studies at different time intervals. The total bacterial cells were obtained using the haemocytometer (direct count) and nutrient agar plates (total plate count). The direct bacterial count showed the maximum of 37.1 x 10 6 CFU/ml at 48th hour whereas by the total plate count the maximum bacterial count of only 21 x 106 CFU/ml was observed at 48th hour. The growth patterns of *A.hydrophila* by the direct count and total plate count methods are presented in Figure 5. The peak growth was observed during 48th hour of culture at room temperature. Figure 6 shows the reisolate count A.hydrophila from the different organs and body fluids of the fish Cyprinus carpio.



Figure 1: Percentage composition of bacterial genera observed in kidney of diseased fish



Figure 2: Percentage composition of bacterial genera observed in liver of diseased fish







Figure 4: Percentage composition of bacterial genera observed in the intestine of diseased fish



Figure 5: Growth of Aeromonas hydrophila isolates at different time intervals



Figure 6: Reisolation of Aeromonas hydrophila from different organs and body fluids

# Determination of LD<sub>50</sub> Live cells of A.hydrophila

The lethal dose (LD<sub>50</sub>) value of *A.hydrophila* live cells for *Cyprinus carpio* was calculated from the probit chart and the percentage mortality for different cell densities is given in Table 2. The LD<sub>50</sub> value was calculated as 3 x 10<sup>6</sup> cells for 16 gm average weight of experimental fishes.

#### DISCUSSION

Most of the bacterial fish diseases reported from many Asian countries are contributing major share for the aquaculture production [21]. Bacteria are considered as serious pathogens, which cause diseases and subsequent economic losses in aquaculture. Among the bacterial diseases, the fishes were predominantly affected by Gram-negative bacteria such as Aeromonas, Pseudomonas and Vibrio and among the three Aeromonas sp. are the main pathogenic organism for the fresh water fishes [22]. Carps are the group of fishes which were used as a research model organism considered as a main source of protein rich food. Species from Aeromonas and Pseudomonas are widely isolated bacterial strains from the carps and its culture environment [23]. A. hydrophila is a causative agent for many diseases for the aquatic organisms and it is major organism investigated by many researchers [24]. Three Indian major

carps include *Catla catla, Labeo rohita, Cirrhinus mrigala* and exotic species like *Cyprinus carpio,* Hypophthalmichthys molitrix and *Ctenopharyngodon idella* are some of the carps tested with *Aeromonas* species. This infection may cause diseases in human beings also through fishes in the food chain [25-28].

## Isolation of pathogenic bacteria from diseased Cyprinus carpio

Based on the morphological, biochemical and physiological characteristics, the isolates were compared with literature reports. The isolated colonies from the diseased Cyprinus carpio were identified as A.hydrophila, Pseudomonas sp., Enterobacter sp and Serratia sp. Among the bacterial isolates A.hydrophila was found to be predominant. Lio Po et al., (1992) [16] recovered bacterial isolates from the fish sample, culture water, soil and the golden snail and reported 89% of the total isolates were the A.hydrophila, 6% of Enterobacter sp and 5% of Pseudomonas sp. Munn and Trust (1984) [20] also reported Aeromonas salmonicida as the primary causative agent of furunculosis affecting mostly the Salmonoids in both fresh water and sea water. Samples from water, fishes, food stuffs and environment are tested for the presence of motile Aeromonas species. Stress may be occurred to the fishes when cultured in polluted water which is affected by the pathogens of Aeromonas species [29,30] and

this can be considered to be a potential pathogens affecting carps [21].

The present investigation is similar to the observations of Areerat et al. [31], Roberts et al. [13] and Liu et.al., [32] who have identified acute infections and abdominal dropsy as the most common syndrome of *Aeromonas* infected *C.carpio*. Amin et al., [2] reported that *A.hydrophila* is frequently associated with diseases in carps, eels, milk fish, channel catfish, tilapia and etroplus. *A.hydrophila* is a highly affecting species which increases mortality rate in a variety of fishes [33,34] and causes enormous economic loss [35].

Previous studies of the bacterial microflora of some fresh water fishes in tropical water showed that Aeromonas species were the most predominant microorganisms isolated from the skin and gills of the fish [36,37]. The same organism has also been isolated from healthy and moribund fishes [38]. Maya et.al, [39] that interaction concluded the of microorganisms with aquatic biota is unique and diverse. When fish are under stress due to the surrounding aquatic environment, saprophytic microorganisms on the skin, gills and in the alimentary tract will turn into pathogen. A.hydrophila has been consistently associated with EUS in fish and the pathogenicity of this bacterium for EUS susceptible fish had been reported [40-42]. A. hydrophila induced severe dermo-muscular necrotic lesions in both catfish and snakehead.

#### **Determination of LD**<sub>50</sub>

In order to find out the suitable root of administration for development of effective management strategies, the intraperitoneal injection was chosen for the inoculation of *A.hydrophila* at  $3x10^6$  cells. Similar type of scientific approach was also made by Irionto and Austin (2002) [43], who have reported the minimal lethal dose of rainbow trout as  $10^5$  cells per fish.

Khalil and Mansour [44] and Daskalov [25] reported that *A.hydrophila* was found to produce haemolytic and proteolytic exotoxin that are lethal to tilapia and the LD<sub>50</sub> value was 2.1 x 10<sup>4</sup> cells/fish. The lethal effect was attributed to the stable unknown virulent factors that were responsible for 20% mortality. Other pathogenic isolates of fish including *Pseudomonas aeruginosa* and *A.hydrophila* were also tested for their pathogenicity [45]. He also observed that the fish isolates of *P. aeruginosa* 

had a lethal dose of  $1.5 \times 10^5$  cells/fish of (*C.carpio*) and  $4.2 \times 10^5$  cells/fish for (*Oreochromis mossambicus*). The fish pathogen *A.hydrophila* had lethal doses of  $2.1 \times 10^6$ ,  $6.8 \times 10^5$  and  $3.2 \times 10^6$  cells/ fish respectively for *C.carpio*, *Labeo rohita*, and *O.mossambicus*.

Lio-Po et al., [46] and Rashid et al., [47] reported that A. hydrophila a minimum dose of 10<sup>6</sup> cfu/ml injected intramuscularly was required to induce dermal lesions in walking catfish. Similarly, Supriyadi [48] showed that A. hydrophila was the most virulent pathogen to walking catfish, and was slightly virulent to giant gourami Osphronemus gourami, and avirulent to common carp *C.carpio* at the dose of 10<sup>5</sup> cells/fish. Most of the bacterial fish diseases were reported from temperate regions. Bacteria are considered as serious pathogens, which cause diseases and subsequent economic losses in aquaculture. Among the bacterial diseases, the fishes were predominantly affected by Gram-negative bacteria such as Aeromonas, Pseudomonas and Vibrio and among the three Aeromonas sp. are the main pathogenic organism for the fresh water fishes [22, 49].

#### CONCLUSION

Water pollution causes acute loss of aquatic production by its excessive load of pathogenic bacteria like Aeromonas hvdrophila. Identification of the water-borne diseases to the protein rich fishes which are considered to be main source of food caters protein supplement to the population in and around the water bodies. In the present study, various pathological bacteria like Aeromonas hydrophila, Pseudomonas species, Serratia marcescens and enterobacteriace species were isolated. Among the bacterial strains Aeromonas hydrophila was found to be the predominant species selected for the growth studies. LD<sub>50</sub>value was calculated to find the potential virulence of the *Aeromonas* hydrophila on Cyprinus carpio, common carp. The present observation showed better understanding of the pathogenic bacteria recorded in the polluted water and created a better option to produce quality aquaculture production for fulfilling the protein need of the society.

#### REFERENCES

[1] Hazen, T.C., Filermans, C. B., Hirsch, R.P., Esch, G. W. 1978. Prevalence and distribution of *Aeromonas hydrophila* in

E-ISSN: 2349 5359; P-ISSN: 2454-9967

the United States. Applied and Environ. Biology. 36.731-738.

- [2] Amin, N. E., Abdulla, I. S., Elallawy, T., Ahmed, S.M. 1985. Motile Aeromonas scepticaemia among Tilapia nilotica (Sarotherodon niloticus) in upper Egypt. Fish Pathology. 20: 93-97.
- [3] Bagarinao, T. 1994. Systematics, distribution, genetics and life history of milkfish, Chanos chanos. Environmental Biology of Fishes, 39: 23-41.
- [4] Azad, I.S., Mandel,B.K., Singh,H.B.K.1992. Aeromonas hydrophila and pathogenicbacteria on diseased Osteobrama balangeri (Val.) from Manipur. Ind. J of Hill. Farmg. 5(1): 59-60.
- [5] Miyazaki, T., Jo ,Y. 1985. A histopathological study of motile aeromonads disease in ayu, *Plecoglossus altevelis*. Fish pathology. 20, 55-60.
- [6] Semel, J. D., Trenholme, G. 1990. Aeromonas hydrophila water associated traumatic wound infections- A review. J Trauma. 30,324-327.
- [7] Altwegg, M., Geiss, H.K. 1989. *Aeromonas* as a human pathogen. CRC. Crit. Rev. Microbiol. 60, 253-287.
- [8] Santosh, Y. I., Bandin, S., Nunez, M., Doranzo, A. E. 1992. Comparitive of the extracellularbiological activities of Vibrio angularium and Aeromonas hydrophila. *Aquaculture* .109, 259-270.
- [9] Neves ,M. S., Nunes, M.P., Millomem, A. M. 1994. *Aeromonas* sps. Exhibit aggregative adherence to Hep-2-cells. J Clin. Microbiol. 32, 1130-1131.
- [10] S. Merino., J. M. Tomás, 2010. Bacterial capsules and evasion of immune responses, in Encyclopedia of Life Sciences, John Wiley & Sons, New York, NY, USA, 3rd edition, 2010,
- [11] Austin, B., D. A. Austin, I. Dalsgaard, B. K. Gudmondsdottir, S. Hoie, J. M. Thornton, J. L. Larsen, B. O'Hici, Powell, R. 1998. Characterization of atypical Aeromonas salmonciida by different methods. Systematic and Applied Microbiology. 21: 50 – 64.
- [12] H. W. Huizinga, G.W. Esch, T. C. Hazen. Histopathology of red-sore disease (*Aeromonas hydrophila*) in naturally and experimentally infected largemouth bass Micropterus salmoides (Lacepede). Journal of fish diseases. 2 (4): 1979:263-277.

- [13] Roberts, R.J., Frerichs, G.N., Miller, S.D., 1992. Epizootic ulcerative syndrome; the current position. *In* : M. Shariff, R.P., Sabasinghe and J.R. Arthur (Eds), *Diseases in Asian Aquaculture - I. Fish Health Section. Asian Fisheries Society*, Manilla, p. 431-436.
- Bejerano,Y., Sarig, S., Roberts, R.J., 1976. Mass mortalities in silver carp, *Hypophthalmichithys morlitrix* (Valenciennes) associated with bacterial infection following handling. J.of Fish. Diseases. 2 49-56
- [15] Lio-Po,G.D., Duremdez-Fernandez, R., 1998. The pathogenicity of bacteria associated with transport-stressed *Chanos chanos* fingerlings. In:*The First Asian Forum*. J.L Maclean, L.V. Hosillos (eds), 227-230. Asian Fisheries Society, Manila. Philippines.
- [16] Lio-Po, G.D., Albright, L.J., Alapide Tendencia. E.V.. 1992. Aeromonas hydrophila in the epizootic ulcerative syndrome (EUS) of snakehead, Ophicephalus striatus and cat fish, Clarius batrachus: quantitative estimation in natural infection and experimental induction of dermomuscular necrotic lesion. In: Diseases in Asian Aquaculture I.M. Shariff, R.P. Subasinghe and J.R. Arthur (eds). Pp 461-474. Fish Health section, Asian Fisheries Society, Manila, Philippines.
- [17] Gopalakrishnan, V.; 1961: Observation on infectious dropsy of Indian major carps and experimental induction. J. Sci. Ind. Res., 20: 357-358.
- [18] Bergey's Manual of Determintative Bacteriology 1998. John G.Holt-Noel.R.Krieg Peter H.A.,Sheath James T.Stanely and T. Williams(Eds.) 9<sup>th</sup> Edn. Lipponcott Williams and Walkins Publications, Philadelphia, PA.19106.USA.
- [19] Lakshmanan, M., Jeyaraman, K., Jeyaraman, J., Ganesan, A. 1971. Laboratory experiments in Microbiology and Molecular biology. Higginbothams Ltd., India 115.
- [20] Reed, L.J. and Muench, H. 1938. A Simple Method of Estimating Fifty Percent Endpoints. American Journal of Hygiene, 27, 493-497.
- [21] Iddya Karunasagar., Indrani Karunasagar., Subhendu Kumar Otta. 2003. Disease Problems Affecting Fish in Tropical

E-ISSN: 2349 5359; P-ISSN: 2454-9967

Environments, Journal of Applied Aquaculture, 13:3-4, 231-249

- [22] Munn, C.B., Trust, T.J., 1984. Role of additional protein in virulence of *Aeromonas salmonicida*, In: *Aquigrup* (Edn). "Fish diseases, Fourth Copraq session" Editora, ATP, Madrid, Spain. 69-79.
- [23] Sanyal, K.B., Mukherjee, D., Guchhait, A., Dash, G.2018. Phenotypic and molecular identification of bacterial species in Indian major carps and exotic carps from south 24 Parganas, West Bengal, India, Int. J. Curr. Microbiol. Appl. Sci 7 (1) 534– 547,
- [24] Gracey, M., Burke, V., Robinson, J. 1982. Aeromonas-associated gastroenteritis, Lancet, 2 (8311) 1304–1306,
- [25] Daskalov, H.2006. The importance of Aeromonas hydrophila in food safety, Food Control 17 (6) (2006) 474–483,
- [26] Ghenghesh, KS., Ahmed, SF., El-Khalek, RA., Al-Gendy A., Klena J. 2008. *Aeromonas* infections in developing countries. Journal of Infection in Developing Countries. 2(2):81–98.
- [27] Kumari, S., Tyor, A.K., Bhatnagar, A. 2019.Evaluation of the antibacterial activity of skin mucus of three carp species. Int Aquat Res 11, 225–239.
- [28] Ramasamy Harikrishnan., Gunapathy Devi., Hien Van Doan., Sundaram Jawahar., Chellam Balasundaram., Kaliyaperumal Saravanan., Jesu Arockiaraj., Mehdi Soltani, Sanchai Jaturasitha.2021. Study on antioxidant potential, immunological response, and inflammatory cytokines induction of glycyrrhizic acid (GA) in silver carp against vibriosis, Fish Shellfish Immunol, 119,193.
- [29] R. Beaz-Hidalgo, A., Martínez-Murcia, M.J., Figueras. 2013. Reclassification of Aeromonas hydrophila subsp. Dhakensis Huys et al. 2002 and Aeromonas aquariorum Martinez-Murcia et al. 2008 as Aeromonas dhakensis sp. nov. comb nov. and emendation of the species Aeromonas hydrophila, Syst. Appl. Microbiol. 36 (3) 171–176.
- [30] Karvonen, A., Rintamaki, P., Jokela, J., Valtonen, E.T.2010. Increasing water temperature and disease risks in aquatic systems, climate change increases the risk of some, but not all, diseases, Int. J. Parasitol. 40 (13) 1483–1488.

- [31] Areerat, S., 1987. Clarias culture in Thailand. Aquaculture, 63: 355-362.
- [32] Liu J, Xie L, Zhao D, Yang T, Hu Y, Sun Z, Yu X. 2019. A fatal diarrhoea outbreak in farm-raised Deinagkistrodon acutus in China is newly linked to potentially zoonotic Aeromonas hydrophila. Transbound Emerg Dis 66:287–298.
- [33] Dias, M.K., Sampaio, L.S., Proietti-Junior, A.A., Yoshioka., E.T., Rodrigues, D.P., Rodriguez, A.F., Ribeiro, R.A., Faria, F.S., Ozório, R.O., Tavares-Dias, M. 2016. Lethal dose and clinical signs of Aeromonas hydrophila in Arapaima gigas (Arapaimidae), the giant fish from Amazon. Vet. Microbiol. 188, 12–15.
- [34] Abdel-Latif, H.M., Khafaga, A.F.2020. Natural co-infection of cultured Nile tilapia Oreochromis niloticus with Aeromonas hydrophila and Gyrodactylus cichlidarum experiencing high mortality during summer. Aquac. Res. 51, 1880– 1892.
- [35] Kumar, R., Pande, V., Singh, L., Sharma, L., Saxena, N., Thakuria, D., Singh, A.K., Sahoo, P.K.2016.Pathological findings of experimental Aeromonas hydrophila infection in golden mahseer (Tor putitora). Fish Aquac. J. 7, 160.
- [36] Chen F., Sun J., Han Z., Yang X., Xian JA., Lv A., Hu X., Shi H.2019. Isolation, Identification and Characteristics of Aeromonas veronii From Diseased Crucian Carp (Carassius auratus gibelio). Front Microbiol. 2019 Nov 26;10:2742.
- [37] Kaper, J.B., Lockman, H., Colwell, R.; Joseph, S.W., 1981: Aeromonas hydrophila. Ecology and Toxigenicity of isolates from an estuary. J Applied Bacteriology. 50, 359-377.
- [38] Praveen Kumar Praveen., Chanchal Debnath, Shashank Shekhar., Nirupama Dalai., Subha Ganguly.2016. Incidence of Aeromonas spp. infection in fish and chicken meat and its related public health hazards: A review. Vet World. Jan; 9(1): 6–11.
- [39] Maya,K., Dhevandaran, K., Natarajan ,P. 1995. Arylsulfatase producing bacteria in the gut of *Therapon jarbua*. International Colloquium on Microbiology in Poikilotherms (ed.R.Lessel), Elsevier Science Publishers, Biochemical Division, 203-210.
- [40] Pathiratne, A., Widanapathirana, G.S., Chandrakanthi,W.H.S.,1994. Association

of *Aeromonas hydrophila* with epizootic ulcerative syndrome (EUS) of freshwater fish in Sri Lanka. J.Appl.Ichthyol. 10, 204-208

- [41] Muthu Ramakrishnan, C., Haniffa, M.A., Jeya Sheela P.2015. Isolation and identification of microbial flora from EUS infected singhi Heteropneustes fossilis. International Journal of Fisheries and Aquatic Studies, 2(4): 178-183.
- [42] Mishra, SS., Das R., Dhiman, M., Choudhary, P., Debbarma, J. 2017. Present Status of Fish Disease Management in Freshwater Aquaculture in India: State-ofthe-Art-Review. J Aquac Fisheries 1: 003.
- [43] Irianto, A., Austin, B., 2002: Use of probiotics to control furunculosis in rainbow trout. Oncorhynchus mykiss (Walbaum). Journal of Fish Diseases. 25: 1-10.
- [44] Khalil, AH., Mansour, EH.1997.Toxicity of crude extracellular product of *Aeromonas hydrophila* in tilapia, *Tilapia nilotica*. Lett Appl Microbiol. 25:269–273.

- [45] Lipton, A.P.; Lakshmanan, M., 1986.
   Microbial diseases of the freshwater fishes of India. Indian Rev. Life Sci. 6: 141 -161
- [46] Lio-Po,G.D., Duremdez-Fernandez, R., 1998. The pathogenicity of bacteria associated with transport-stressed *Chanos chanos* fingerlings. In:*The First Asian Forum*. J.L Maclean, L.V. Hosillos (eds), 227-230. Asian Fisheries Society, Manila. Philippines.
- [47] Rashid M.M., Hossain M.S., Ali M.F.2013. Isolation and identification of *Aeromonas hydrophila* from silver carp and its culture environment from Mymensingh region. J. Bangladesh Agric. Univ. 11(2):373–376.
- [48] Supriyadi,H., 1986: The susceptibility of various fish species to infection by the bacterium *Aeromonas hydrophila*. In: *The First Asian Fisheries Forum*. J.L.Maclean, L.B. and Diazon and L.V.Hosillos (eds.), pp241-242. Asian Fisheries Society, Manila, Philippines.
- [49] Johnson, P.J., Paull, S.H. 2011.The ecology and emergence of diseases in freshwaters. Freshwater Biol. 56:638– 657.

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